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Bioprocess-Inspired Synthesis

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Mussel-Directed Synthesis of Nitrogen-Doped Anatase TiO₂

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Abstract: Structure-forming processes leading to biominerals are well worth learning in pursuit of new synthetic techniques. Strategies that attempt to mimic nature in vitro cannot replace an entire complex natural organism, requiring ingenuity beyond chemists' hands. A "bioprocess-inspired synthesis" is demonstrated for fabrication of N-doped TiO2 materials at ambient temperature by direct implantation of precursor into living mussels. The amorphous precursor transforms into Ndoped anatase TiO2 with a hierarchical nanostructure. Synthetic TiO₂ exhibits high phase stability and enhanced visiblelight photocatalytic activity as a result of modifications to its band gap during in vivo mineralization. Intracellular proteins were found to be involved in TiO₂ mineralization. Our findings may inspire material production by new synthetic techniques, especially under environmentally benign conditions.

In studying and mimicking the well-defined structures or unique functions of biomaterials, [1-3] scientists have succeeded in designing and synthesizing bio-inspired materials or bioinspired functions. [4-6] Furthermore, in contrast with anthropogenic technologies that commonly require harsh conditions, the fantastic structure-forming processes found in biological systems are the results of many billions of years of evolution, leading to efficient and accurate fabrication of biominerals under environmentally benign conditions.^[7,8] Hence, natural structure-forming processes themselves are also worth understanding when developing new synthesis techniques for materials. This field of research is referred to as "bioprocess-inspired fabrication". To date, bioprocessinspired work focuses on using organic matrices, including proteins, as templates for directing the formation of inorganic materials in vitro. [9-11] However, in most situations, a single protein or a combination of several proteins cannot be substituted for an entire natural organism, which is a capable cradle for the growth of biominerals (formation of a pearl inside a mussel for example). The complexity and ingenuity of biological systems are still beyond the skills of chemists to recreate. [12,13] Herein, we propose a strategy that capitalizes on natural organisms as platforms for fabricating inorganic materials at ambient temperature. A significant challenge remains as to whether we can directly use biosynthetic pathways in living organisms without genetic modification, to deliberately produce anthropogenically useful materials inessential to those organisms.

The mussel is a typical bio-mineralization system for pearl formation. Among them, the fresh-water mussel Cristaria plicata is extensively used for culturing blister pearls of large sizes and various shapes, and is therefore a beneficial host for preparation of samples (Figure 1a). Herein, we use Cristaria plicata as a cradle for synthesizing TiO₂, a material with broad applications to facilitate solving both energy and environmental problems.[14-17] To improve its energy conversion efficiency in the visible light regime, persistent efforts have been devoted to modifying its inherent band gap. In particular, nitrogen-doped TiO2 (N-TiO2) appears to use visible light efficiently, but heat treatments and complicated procedures are usually required during preparation.^[17,18] A one-pot synthetic method for tailoring key properties of TiO₂ (such as microstructure, phase stability and light-use efficiency), especially at ambient temperature, is of great significance from an energy-saving perspective.^[19]

Based on a natural pearl culture process, we demonstrate a novel "one-pot" ambient-temperature approach for fabrication of N-TiO₂ materials using living Cristaria plicata. Interactions between the organism and TiO₂ precursor give rise to compelling questions regarding bio-inspired synthesis, such as: the influence of the host upon chemical reactions governing formation, the structures obtainable, and the resultant properties of products. Our results show that amorphous precursor transforms into N-doped anatase TiO₂ with hierarchical nanostructure when grown inside living mussels. Synthetic TiO₂ exhibits enhanced visible-light photocatalytic activity in both chemical waste remediation and hydrogen generation.

Amorphous TiO₂ precursor was shaped into a tablet with a diameter of 10 mm and thickness of 1-2 mm, and then implanted as a pearl nucleus between the mantle and shell of Cristaria plicata (Supporting Information, Scheme S1). TiO₂ precursor was prepared using a typical sol-gel method, by adding 1 mL of acetic acid to 5 mL of tetrabutyl titanate. The precursor was an amorphous phase (Figure 1 b,c), even after

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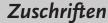
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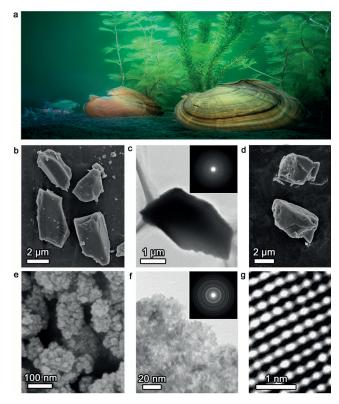


Figure 1. a) Living *Cristaria plicata*. b) SEM and c) TEM images of TiO_2 precursor. Inset: the SAED pattern. d) SEM image of TiO_2 precursor aged in air for 120 days. e) SEM image of TiO_2 nanoparticles mineralized in mussel for 90 days. f) TEM image of TiO_2 nanoparticles. The diffraction rings of SAED pattern are assigned to the (101), (004), (200), (211), (204), (116), and (215) planes of anatase. g) An HRTEM image of TiO_2 revealing the fringes of (101) planes with a lattice spacing of about 0.33 nm.

aging in air for 120 days (Figure 1 d). However, after mineralization within the mussel's body for 90 days, agglomerates consisting of nanospherical grains were obtained (Figure 1e). The bm-TiO₂ crystallized as shown, with well-resolved rings and lattice features in the SAED and HRTEM images (Figure 1 f,g). X-ray diffraction patterns of bm-TiO₂ present broad diffraction peaks, revealing the nanosized nature of crystallites (Figure 2a). The crystal phase has an anatase structure with an average grain size of approximately 10 nm, in agreement with TEM observations. More detailed information about crystallization processes was obtained by a time-resolved study over the course of mineralization, starting from the earliest stage (Supporting Information, Figure S1). When the precursor was aged in air for 120 days, no sign of crystallization was observed (Supporting Information, Figure S1g). Normally, the amorphous TiO2 obtained from the sol-gel method requires heat treatments before crystallization occurs.[19]

The BET surface area and pore volume of bm-TiO₂ were determined to be $206 \, \mathrm{m^2 g^{-1}}$ and $0.22 \, \mathrm{m^3 g^{-1}}$ respectively (Figure 2b). The isotherm of bm-TiO₂ revealed two hysteresis loops at relative pressure range of 0.45–0.8 and 0.9–1.0, indicating the presence of hierarchical meso/macropores. The mesopore size is centered at 3.8 nm. In contrast, abundant

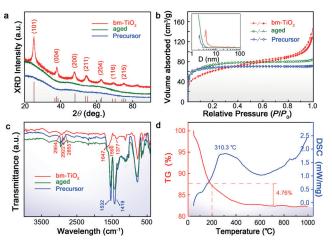


Figure 2. Structure evolution of bm-TiO $_2$ nanocrystals. a) X-ray diffraction analyses of TiO $_2$ samples. b) N $_2$ adsorption-desorption isotherms of TiO $_2$ samples. Relative pore size distributions are shown in the inset. c) FTIR spectra of TiO $_2$ samples. d) Thermogravimetric analysis of a bm-TiO $_2$ sample after mineralization.

micropores were present in the ${\rm TiO_2}$ precursor. After aging in air for 120 days, the precursor isotherm also presented microporous features. During the biomineralization process, micropores convert into hierarchical meso/macropores due to crystallization and grain growth. Therefore, by using living mussels as platforms, hierarchically porous ${\rm TiO_2}$ powders are synthesized at ambient temperature. Studies indicate that hierarchically porous structure can improve photocatalytic activity. [16,20]

A change in chemical composition was found in the bm-TiO₂ sample during mineralization (Figure 2c). Precursor prepared by adding acetic acid exhibits characteristic C=O stretching vibrations at 1532 and 1419 cm⁻¹, variously coordinated to Ti as bidentate ligands, [19] which are stable and usually require post-calcination before elimination. The two bands are still visible in samples after aging in air for 120 days or after heating at 300°C (Supporting Information, Figure S2). Interestingly, after in vivo mineralization, the bands completely disappear, implying that Ti-acetate complexes have been replaced by other species. Furthermore, several substantial bands associated with proteins are observed, corresponding to amide I, II, and III vibrational modes at 1647, 1590, and 1377 cm⁻¹, respectively. [21] To estimate the organic content, we applied a thermogravimetric analysis procedure to the fresh bm-TiO₂ (Figure 2d). Weight loss below 200 °C is probably due to loss of water, and 4.76 % mass loss above 200°C is induced by decomposition of organic matrices.[22]

Pristine TiO_2 was prepared as a reference by directly heating the TiO_2 precursor at 600 °C for 30 min. The results show that pristine TiO_2 has the characteristics of anatase (Supporting Information, Figure S3). A typical single O 1s peak for pristine TiO_2 is observed at 530 eV, whereas for bm- TiO_2 nanocrystals the O 1s peak is resolved into two peaks at 529.8 and 531.8 eV. The 531.8 eV peak is attributed to Ti-OH species (Supporting Information, Figure S4). [14] The broad N 1s peak in bm- TiO_2 can be deconvoluted into two peaks





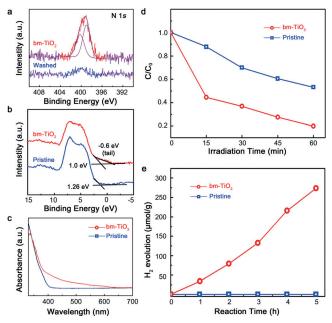


Figure 3. a) N 1s XPS spectra of bm-TiO₂ nanocrystals before and after chemical washing. The red and blue curves are XPS data, and the purple curves are the fitting of experimental data. b) Valence-band XPS spectra of TiO₂ samples. c) UV/Vis absorption spectra of TiO₂ samples. d) Visible-light-promoted photocatalytic degradation of RhB carried out by TiO2 samples. e) Hydrogen evolved under visible light after photocatalytic water reduction using TiO₂ samples.

centered at 400.2 eV and 398.9 eV (Figure 3a). For comparison, we removed residual proteins by washing bm-TiO2 according to a typical chemical method. After washing, the peak at 398.9 eV disappeared but the peak at 400.2 eV remained. Therefore, the peak at 398.9 eV is assigned to N-H bonds arising from residual proteins, and the peak at 400.2 eV is related to interstitial nitrogen dopant.[17,23] The atomic concentration of nitrogen was calculated based on the XPS data, revealing that 0.44 at. % N has been doped into bm-TiO₂. Pristine TiO₂ displays valence band density of states (DOS) characteristic of anatase, with a band edge about 1.26 eV below the Fermi energy. By comparison, the valance band of bm-TiO2 shows differences: the main absorption onset is located at about 1.0 eV and the maximum energy related to the band tail blue-shifts toward the vacuum level at about 0.6 eV (Figure 3b). The band gap shift is attributed to lattice defects such as those arising from interstitial nitrogen.

Natural proteins exhibit improved mineral-forming activities in vivo, as compared to in vitro processes utilizing isolated or synthetic proteins, owing to the cellular control for these natural proteins.^[24] Thus, under these special conditions, self-adding active proteins act as a nitrogen source for N-doping, which could lead to production of TiO₂ with enhanced visible-light photocatalytic activity. Pristine TiO2 is unable to absorb visible radiation owing to its relatively large band gap^[17] (Figure 3c), whereas bm-TiO₂ shows visible light absorption from 380-650 nm owing to interstitial nitrogen doping. bm-TiO₂ exhibits superior photocatalytic activity in degradation of Rhodamine B (RhB) compared with pristine TiO₂ (Figure 3 d). The photocatalytic activity of bm-TiO2 does not significantly decrease even after five cycles of photodegradation, implying that these photocatalysts are very stable (Supporting Information, Figure S5a). Notably, pristine TiO₂ also demonstrates photocatalytic activity under visible light owing to a self-sensitized degradation process.^[25] To eliminate this effect, phenol degradation was also investigated and a similar superior performance was observed in the presence of bm-TiO₂ nanocrystals (Supporting Information, Figure S5b).

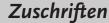
A photocatalytic test for hydrogen evolution was carried out under visible light (Figure 3e). No obvious hydrogen production was observed using pristine TiO2, as expected. Whereas for bm-TiO2 the average rate of hydrogen production increased to 55 µmol h⁻¹g⁻¹, reflecting the activity of nitrogen interstitial gap states.^[14,17] A limited number of reports demonstrate that TiO2 material fabricated at ambient temperature has the ability to produce hydrogen under visible-light irradiation. Thus, it can be concluded that the visible-light efficiency of TiO₂ is improved directly in Cristaria plicata at ambient temperature.

Stability of the anatase phase is also an important factor for TiO₂ photocatalysts. Previous studies indicate that anatase to rutile phase transformation usually occurs at 500–700 °C. [19] In the present work, TiO₂ precursor completely transforms into rutile at 700 °C. In contrast, the temperature required for phase conversion of bm-TiO₂ increases to 1000 °C (Supporting Information, Figure S6), revealing the high phase-stability of this material.

In the present work, hierarchically meso/macroporous N-TiO₂ is obtained in living Cristaria plicata at ambient temperature with excellent properties. Our findings inspire a new synthesis technique to fabricate materials by directly using natural organisms. To support and extend the current research, we also investigated the mineralization of other materials in mussels and obtained numerous interesting results. For example, macroscopic CaCO₃/Fe₂O₃ composite rods were formed by oriented growth of single-crystal CaCO₃ along implanted Fe₂O₃ nanocrystals (Supporting Information, Figure S7a). Oriented MgO material with enhanced absorptive activity was obtained via self-organization and mesoscale transformation (Supporting Information, Figure S7b).

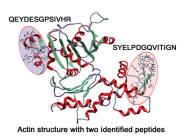
To understand how living organisms direct the formation of these materials, proteins in a bm-TiO2 sample were extracted and analyzed by mass spectrometry. Extracellular matrices were deemed to be involved in the bio-mineralization processes for nacre. [12,13] However, almost all the identified proteins are intracellular proteins. Actin and histone are the most abundant among these, constituting the cytoskeleton and chromatin, respectively (Supporting Information, Table S1; Figure 4). Most identified segments of actin are on the surface and are therefore accessible to TiO₂ (Figure 4; Supporting Information, Figure S8). Ubiquitin, a cytoplasmic protein that regulates the inflammatory responses and programmed cell death, was also identified; thereby implying that cell lysis occurs during mineralization of TiO₂. As reported, [26-28] nanomaterials such as TiO₂ could induce various kinds of cytotoxicity, which cause a stress responses in mussels other than death. Amorphous TiO₂ exhibits enhanced cytotoxicity in comparison with the anatase

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Representative proteins identified by mass spectroscopy

Identified Proteins	Subcellular Location
Actin	Cytoplasm
Histones H ₄ , H ₂ A, H ₂ B	Nucleus
Ubiquitin	Cytoplasm
Glyceraldehyde 3- phosphate dehydrogenase	Cytoplasm
Cu-Zn superoxide dismutase	Cytoplasm

Figure 4. Representatives of the proteins or peptides found in $bm-TiO_2$ nanocrystals.

phase.^[28] Pearl production is usually initiated by implanting an inorganic bead (pearl nucleus). The bead is gradually encapsulated by vast cells such as hemocytes.^[13] In the present case, cytotoxicity of the amorphous TiO₂ precursor acting as a bead may lead to cell rupture. Intracellular proteins released into the bead direct the amorphous-to-anatase transition, thereby lowering the cytotoxicity of TiO₂ toward mussels.^[27,28] In a sense, this phase transition may be physiologically required by the mussel. Additionally, some proteins may further degrade and produce ammonia, phenols and amines in living system, which may also promote nitrogen doping at ambient temperature.^[29]

It is known that the amorphous-to-anatase phase transition is accomplished by structural rearrangement of TiO_6^{2-} octahedral units. The dehydration between octahedra, resulting in shared edges, is the rate-limiting step of this transition.^[30] It is possible that in vivo crystallization of TiO₂ relies on biomolecules to control phase transition by removing H₂O, which also occurs in CaCO₃ biomineralization.^[11] Taking peptides from actin as an example (Supporting Information, Figure S9), we hypothesize that the OH groups of proteins may act as a proton transfer catalyst[31] to facilitate the dehydration. A density functional theory (DFT) study was also performed (Supporting Information, Figure S10). [32-34] It was found that direct dehydration has an energy barrier of 18.5 kcal mol⁻¹. In contrast, OH-catalyzed dehydration adopts a proton-transport catalysis strategy with a low energy barrier of 10.9 kcal mol⁻¹.

We also set up in vitro experiments in three kinds of aqueous solutions (Supporting Information, Figure S11): with actin, water, and extrapallial fluid protein. HRTEM and SEAD clearly indicate that an acceleration of the crystallization process is achieved in actin solution. XRD patterns also show that the crystallization process is faster in actin. However, no nitrogen doping occurs during in vitro mineralization, suggesting that doping is induced by the synergistic effects present in living systems (Supporting Information, Figure S12). Therefore, the detailed mechanism of in vivo

formation is not easily accessible by in vitro experiments. Additional work with further controls on the mineralization process will be carried out in the future.

In conclusion, for the first time, living mussel has been used as a cradle for fabrication of TiO₂ materials (which are not required in its physiological processes) with optimized properties at ambient temperature. Although this technique is still in its infancy, our approach extends present chemical methodologies for material production, especially under environmentally benign conditions.

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- [1] S. Weiner, W. Traub, H. D. Wagner, J. Struct. Biol. 1999, 126, 241.
- [2] J. Aizenberg, A. Tkachenko, S. Weiner, L. Addadi, G. Hendler, Nature 2001, 412, 819.
- [3] A. Veis, Science 2005, 307, 1419.
- [4] E. Munch, M. E. Launey, D. H. Alsem, E. Saiz, A. P. Tomsia, R. O. Ritchie, *Science* 2008, 322, 1516.
- [5] L. Jiang, Y. Zhao, J. Zhai, Angew. Chem. Int. Ed. 2004, 43, 4338;Angew. Chem. 2004, 116, 4438.
- [6] W. L. Noorduin, A. Grinthal, L. Mahadevan, J. Aizenberg, Science 2013, 340, 832.
- [7] S. Mann, *Biomineralization*, Oxford University Press, Oxford,
- [8] W. D. Kingery, Introduction to ceramics, Wiley, New York, 1960.
- [9] S. L. Sewell, D. W. Wright, Chem. Mater. 2006, 18, 3108.
- [10] C. Du, G. Falini, S. Fermani, C. Abbort, J. Moradian-Oldak, Science 2005, 307, 1450.
- [11] N. Kröger, M. B. Dickerson, G. Ahmad, Y. Cai, M. S. Haluska, K. H. Sandhage, N. Poulsen, V. C. Sheppard, *Angew. Chem. Int. Ed.* 2006, 45, 7239; *Angew. Chem.* 2006, 118, 7397.
- [12] L. Addadi, D. Joester, F. Nudelman, S. Weiner, *Chem. Eur. J.* 2006, 12, 980.
- [13] M. Awaji, A. Machii, Aqua-Bio Science Monographs 2011, 4, 1.
- [14] X. B. Chen, L. Liu, P. Y. Yu, S. S. Mao, Science 2011, 331, 746.
- [15] G. Q. Zhang, H. B. Wu, T. Song, U. Paik, X. W. Lou, Angew. Chem. Int. Ed. 2014, 53, 12590; Angew. Chem. 2014, 126, 12798.
- [16] J. G. Yu, Y. R. Su, B. Cheng, Adv. Funct. Mater. 2007, 17, 1984.
- [17] J. Wang, D. N. Tafen, J. P. Lewis, Z. L. Hong, A. Manivannan, M. J. Zhi, M. Li, N. Q. Wu, J. Am. Chem. Soc. 2009, 131, 12290.
- [18] R. Asahi, T. Morikawa, T. Ohwaki, K. Aoki, Y. Taga, Science 2001, 293, 269.

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- [19] J. F. Ye, W. Liu, J. G. Cai, S. Chen, X. W. Zhao, H. H. Zhou, L. M. Qi, J. Am. Chem. Soc. 2011, 133, 933.
- [20] Y. Li, Z. Y. Fu, B. L. Su, Adv. Funct. Mater. 2012, 22, 4634.
- [21] P. R. Carey, Biochemical Applications of Raman and Resonance Raman Spectroscopies, Academic Press, New York, 1982, chap. 4.
- [22] Y. J. Lee, H. Yi, W.-J. Kim, K. Kang, D. S. Yun, M. S. Strano, G. Ceder, A. M. Belcher, *Science* 2009, 324, 1051.
- [23] V. Etacheri, M. K. Seery, S. J. Hinder, S. C. Pillai, *Chem. Mater.* 2010, 22, 3843.
- [24] L. B. Gower, Chem. Rev. 2008, 108, 4551.
- [25] W. Zhao, C. C. Chen, W. H. Ma, J. C. Zhao, D. X. Wang, H. Hidaka, N. Serpone, *Chem. Eur. J.* 2003, 9, 3292.
- [26] M. I. Setyawati, C. Y. Tay, Nat. Commun. 2013, 4, 1673.
- [27] Z. Xu, X. W. Liu, Y. S. Ma, H. W. Gao, Environ. Sci. Pollut. Res. 2010, 17, 798.

- [28] I. L. Hsiao, Y. J. Huang, Sci. Total Environ. 2011, 409, 1219.
- [29] C. Burda, Y. L. Lou, X. B. Chen, A. C. S. Samia, J. Stout, J. L. Gole, *Nano Lett.* **2003**, *3*, 1049.
- [30] Y. Kazumichi, O. James, J. Phys. Chem. B 1999, 103, 7781.
- [31] Q. F. Shi, X. Li, Y. Z. Xia, L. M. Zhang, Z. X. Yu, J. Am. Chem. Soc. 2007, 129, 15503.
- [32] C. Lee, T. W. T. Yang, R. G. Parr, Phys. Rev. B 1988, 37, 785.
- [33] M. J. Frisch, Gaussian 09, revision A.02; Gaussian, Inc.: Wallingford, CT, 2009, 785.
- [34] K. Fukui, Acc. Chem. Res. 1981, 14, 363.

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